

The challenge of treating central nervous system infections in the developing world

Ranawaka U K¹

Journal of the Ceylon College of Physicians, 2018, 49, 2-15

Introduction

Central nervous system (CNS) infections produce high rates of morbidity and mortality. Case fatality rates are in the range of 17-33% in bacterial meningitis¹⁻³, 13-65% in tuberculous meningitis (TBM)^{4,5}, and 20-30% in Japanese encephalitis (JE)^{6,7}. Residual neurological sequelae are common among survivors; 30-54% after bacterial meningitis^{3,8}, 20-47% after TBM^{4,5}, and 50-60% after JE^{6,7}. CNS infections produce greater challenges to physicians in developing countries like Sri Lanka, as they are commoner and produce more deaths and disability in these regions⁹⁻¹³. Delays in treatment are associated with excess mortality and residual neurological deficits, whereas early treatment has been shown to improve outcome^{2,7,10,14-17}.

Early and appropriate treatment depends on rapid and accurate diagnosis. Predicting the type of CNS infection based on clinical findings alone, however, is difficult and unreliable¹⁸. Typical symptoms of meningitis were shown to have low sensitivity for diagnosis in a meta-analysis¹⁹. The classical triad of fever, neck stiffness and altered mental state is seen in only 44-46%^{1,3,19,20}. Traditional signs of meningeal irritation are poor predictors of meningitis, with a sensitivity of 30% for neck stiffness, and only 5% for Kernig's sign and Brudzinski's sign²¹. Establishing the diagnosis, therefore, depends on laboratory investigations, especially cerebrospinal fluid (CSF) analysis. CSF parameters that can independently predict bacterial meningitis (CSF: blood glucose ratio <0.23, protein >2.2g/L, leucocyte count >2000/cc, and a polymorphonuclear leucocyte count >1180/cc) are well recognised²². However, such changes reflect more severe infection, and are seen only in a minority of cases. CSF cytology, which is widely used in early decision-making, can be negative in up to 10% with bacterial meningitis, and importantly, these patients have poorer outcomes^{20,23}. Simple microbiological

staining measures that can guide early treatment have poor diagnostic yields. Gram staining of CSF has good specificity in bacterial meningitis (up to 97%), but can be negative in 40-75%²⁴. In TBM, sensitivity of Ziehl-Nielsen staining for acid-fast bacilli (AFB) is only 5-30%²⁵.

Targeted treatment therefore requires isolation of the organisms in blood or CSF, but this is often hindered in developing countries by inadequate diagnostic facilities^{10,26}. Blood cultures are positive in about two-thirds of patients with bacterial meningitis in developed countries^{10,20}, but rates fall to less than one-third in the developing world^{10,27,28}. CSF isolation rates are high (60-73%) in developed countries^{1,20,22}, but much lower (3-35%) in developing countries^{10,29-32}. Tests such as polymerase chain reaction (PCR) based assays for nucleic acid detection facilitate rapid and accurate diagnosis³³; they are, however, not readily available in developing countries. Several other factors related to resource constraints, such as the lack of treatment facilities, trained personnel and effective treatments adversely affect management of CNS infections in these settings. Clinical decisions on presumptive diagnosis and empiric treatment can only be guided by knowledge of local disease patterns, but such epidemiological data are often lacking from developing countries.

In Sri Lanka, data on the aetiology of CNS infections is limited. Previous published work has been largely confined to individual case reports or case series of specific syndromes^{29,30,34-37}.

Methodology

This oration is based on data from two prospective observational studies designed to describe the clinical spectrum, accuracy of diagnosis, management and early outcome of CNS infection at a Sri Lankan tertiary care hospital (Colombo North Teaching Hospital, Ragama – CNTH). These are the first prospective studies involving unselected adult patients with presumed CNS infection in Sri Lanka.

¹ Professor in Neurology, Department of Medicine, Faculty of Medicine, University of Kelaniya, Sri Lanka.



Study 1 enrolled all adult patients with suspected CNS infection attending all the medical wards of the CNTH. It was conducted over a two-year period (July 2007 to June 2009), and studied 215 adult patients³⁸.

Study 2 enrolled all adult and paediatric patients with suspected CNS infection attending all the medical and paediatric wards of the CNTH, and was conducted over a similar duration (July 2009 to June 2011), and studied 374 patients (212 adults, 162 children)^{39,40}. In an extension to Study 2, we aimed to improve the diagnostic yield by collaborating with an overseas institution for analysis of CSF samples in a reference laboratory (Oita University, Japan). The two studies employed similar inclusion criteria, and diagnostic criteria to classify CNS infection. Criteria for enrolment were as follows: any combination of the triad of fever, headache, and vomiting, with any of the following: altered level of consciousness, new-onset seizures, focal neurologic deficits, altered behavior, and signs of meningeal irritation³⁸.

Data collection

We collected data regarding demographic and clinical characteristics, laboratory findings, aetiologies detected, treatment given and early outcome. Patients had standard clinical evaluation, diagnostic work up including neuroimaging and CSF analysis, and treatment available at the CNTH. Management decisions were made by the attending consultants. No additional investigational procedures were performed or treatments given for the purpose of the study. Data was collected by trained pre-intern medical officers who visited the wards every day of the year (including holidays) to ensure completeness of data. All patients were seen within 24 hours of admission and reviewed daily until discharge/death. Information was obtained by interview of patients and relatives, and supplemented by case record review. The data collection instruments were pre-tested in a pilot study over a 3-month period (April-June 2007). Data was entered into a computerised database and analysed using standard statistical software.

Investigation for aetiology

Blood investigations included total and differential leucocyte counts, erythrocyte sedimentation rate (ESR), C-reactive protein (CRP) levels and bacterial culture. CSF was subjected to protein and glucose measurement, total and differential leucocyte counts, Gram staining, staining for acid-fast bacilli (AFB) and bacterial culture.

CSF samples from patients admitted during the period July 2009-November 2010 were sent to the Oita University, Japan for detailed analysis (**Study 2-Extension**). Bacteria were tested by PCR amplification of 16S rRNA, followed by sequencing. The following viruses were tested for by PCR: herpes simplex virus (HSV)-1, HSV-2, varicella-zoster virus (HSV-3), Epstein-Barr virus (human herpesvirus [HHV] type 4), cytomegalovirus (HHV-5), HHV-6, HHV-7, HHV-8, dengue virus, Japanese encephalitis virus, rubella virus, West Nile virus, yellow fever virus, tick-borne encephalitis virus, Nipah virus, measles virus, mumps virus, parainfluenza virus, respiratory syncytial virus, metapneumovirus, Chikungunya virus, Sindbis virus, Semliki Forest virus, eastern equine encephalitis virus, western equine encephalitis virus, poliovirus, Coxsackie virus, echovirus, enterovirus, lyssaviruses, Chandipura virus, bocavirus (HBoV), rotavirus, astrovirus, norovirus, parechovirus, and human adenovirus (HAdV). On-cell Western analysis (a cell-based assay) was done for anti-N-methyl-D-aspartate receptor (NMDAR) antibodies. CSF samples negative for an aetiological agent were further analysed by deep sequencing for viral genome detection^{39,40}.

Case definitions and diagnostic classification

The initial presumptive diagnosis and the final diagnosis entered at the time of discharge were recorded. The clinical syndrome of CNS infection was classified as meningitis (bacterial, tuberculous or viral), encephalitis, 'meningo-encephalitis', cerebral malaria and cerebral abscess. Data was reviewed after discharge (or death) to assess the accuracy of diagnosis, and the proportion of patients in whom a definitive aetiological diagnosis was established. The final diagnosis of CNS infection was categorized as 'definite', 'probable', 'possible' and 'uncertain', based on the strength of association between clinical features and investigation findings (blood and CSF analysis, neuroimaging, EEG). Patients in whom a diagnosis other than CNS infection was established were labeled as 'alternate diagnosis'³⁸. (Table 1) These categories were based on previously used criteria^{27,41,42,43,44}.

Ethical considerations

Ethical approval for both studies was granted by the Ethics Review Committee of the Faculty of Medicine, University of Kelaniya. The extension to Study 2 was also approved by the Ethics Committee of the Oita University, Japan. Informed written consent was obtained from all participants, or from a relative/guardian when the participant was unable to provide consent. Administrative approval was obtained from the Director, CNTH.

Results

Demographic and clinical characteristics

The two studies provide data on 589 consecutive patients with CNS infections (427 adults, 162 children) admitted to a single centre over a 4-year period. The main findings are summarised in Tables 2-6. The mean age was 43.1 years (± 19.7) in adults, and 3.1 years (± 3.1) in children. Males predominated among

adults (57.8%), but not in children (42.6%) ($p=0.001$) (Table 2). The classical clinical features such as fever, headache, vomiting and altered consciousness were commonly seen. Headache, altered consciousness, focal signs and meningism were commoner among adults ($p<0.001$), whereas seizures and behavioural changes were commoner among children ($p<0.001$). Prior antibiotic use was common (25.3%) (Table 2).

Table 1. Case definitions and diagnostic classification of CNS infection^{27,41,42,43,44}

1. On case note review after discharge/ death, the diagnosis in each case was classified as:

Diagnosis Uncertain	<ul style="list-style-type: none"> compatible clinical picture with or without supporting blood results CSF cytology not available EEG findings or imaging findings not available no confirmation of aetiology (CSF Gram stain, blood or CSF culture, CSF antigen test, blood or CSF serology or PCR)
Alternative diagnosis	CNS infection excluded – <ul style="list-style-type: none"> compatible clinical picture with or without supporting blood result but, negative CSF findings and negative EEG/ imaging findings OR alternative diagnosis found
Possible diagnosis	<ul style="list-style-type: none"> compatible clinical picture with or without supporting blood results CSF cytology normal no supportive evidence on EEG or imaging no confirmation of aetiology (CSF Gram stain, blood or CSF culture, CSF antigen test, blood or CSF serology or PCR) no alternative diagnosis found
Probable diagnosis	<ul style="list-style-type: none"> compatible clinical picture supporting CSF cytology or EEG or imaging findings no confirmation of aetiology (CSF Gram stain, blood or CSF culture, CSF antigen test, blood or CSF serology or PCR) no alternative diagnosis found
Definite diagnosis	<ul style="list-style-type: none"> clinical evidence AND positive organism detection (CSF Gram stain, blood or CSF culture, CSF antigen test, blood or CSF serology or PCR result) no alternative diagnosis found

(Continued)

(Table 1 continued)

2. Infection syndromes were classified as:

Meningitis (bacterial, tuberculous or viral)	fever, headache, vomiting, meningeal signs
Encephalitis	fever, headache, vomiting, altered level of consciousness, seizures, focal neurological deficits, altered behavior, no meningeal signs
Meningo-encephalitis	mixed picture – both meningitic and encephalitic features
Cerebral abscess	<ul style="list-style-type: none"> • Meningo-encephalitis PLUS focal neurological deficits • abscess noted on CT/MRI
Cerebral malaria	blood film positive for <i>P. falciparum</i>

3. Classification of investigation results**Blood results were classified as:**

Suggestive of bacterial infection	raised white cell count (WCC) >11,000/mm ³ , and elevated neutrophils >60%, elevated ESR or CRP
Suggestive of viral infection	low or normal WCC and elevated lymphocytes >50%

CSF results were classified as:

Suggestive of bacterial infection	turbid or cloudy, elevated proteins, low glucose (low CSF: blood glucose ratio – <50%), elevated WBC >100/mm ³ , elevated neutrophils >60%
Suggestive of viral infection	clear, normal glucose, normal or elevated protein (>45 mg/dl), elevated WBC >5/mm ³ , high mononuclear cell/lymphocyte count- >50%
Suggestive of tuberculous infection / partially treated bacterial infection	turbid, elevated protein, low glucose, elevated WBC, high mononuclear cell/lymphocyte count- >50%
Normal	clear, proteins normal (<45 mg/dl), sugar normal (CSF: blood glucose ratio >50%), cells <5/mm ³ and all lymphocytes

Investigation findings

Peripheral blood white blood cell (WBC) counts were done in over 95% of patients, and were elevated in the majority. In contrast, CRP was tested in less than 50% of patients due to lack of availability at CNTH at the time of the studies; it was elevated in over 50% of those tested. Blood cultures were done in 33.7% of adults and 51.9% of children. In Study 1, a positive blood culture was seen in one patient (*S. pneumoniae*). In Study 2, blood cultures were positive in five children (*S. pneumoniae*=1, *H. influenzae*=1, *Staph. aureus*=3) and three adults (*S. pneumoniae*=2, *Staph. aureus*=1).

CT scanning was done in 74% of adults, with features suggestive of cerebral oedema in 22.2%; EEG was done in 36.8% adults, and showed changes suggestive of CNS infection in 28%; the respective numbers were much smaller for children (Table 3).

CSF analysis was done in all children, and in 80.2% of adults (overall 81.2%). Findings even remotely suggestive of bacterial infection were seen in only a few patients (low glucose - 13%, presence of at least one neutrophil - 31.1%). CSF changes more likely to indicate bacterial meningitis (cells >100 WBC/mm³;

neutrophils >80% of WBC; ratio of CSF:blood glucose <0.4; protein >50 mg/dl)^{20,45,46} were seen in only four patients. CSF Gram stains, AFB stains and cultures were all negative (Table 3). PCR testing in CSF was done in only a few patients due to lack of availability in the state sector and cost constraints; one was positive for *H. influenzae*, and all other samples tested were negative for bacteria and TB.

With the facilities available at the CNTH, a microbiologically proven 'definitive' diagnosis of CNS infection was established in only 12 (2%) patients (Study 1- one patient, 0.5%; Study 2- eleven patients, 2.9%). According to the diagnostic criteria, a diagnosis of CNS infection was considered 'probable' in 52.6% of patients, and 'possible' in 9%. The syndromic diagnosis (in 'definite', 'probable' and 'possible'

categories) was meningitis in 32.4% of patients, encephalitis in 12.9%, and meningo-encephalitis in 19.2%. Five patients had a cerebral abscess diagnosed on neuroimaging. Diagnosis of CNS infection was considered 'uncertain' in 13.2%, and an 'alternative diagnosis' was found in 21.4% (Tables 4,6).

Treatment and Outcome

Over 90% of the patients were given intravenous antimicrobial therapy on empiric grounds, mostly without microbiological confirmation; 3rd generation cephalosporins in 78.6% of patients, and acyclovir in 56.2%. Intravenous steroids were used in 31.9% of patients. Mean duration of hospital stay (SD) was 8.9 (5.7) days. Most patients (81%) were discharged home, and 31 patients (5.3%, all adults) died during hospital stay (Table 5).

Table 2. Demographic and clinical characteristics

	Study 1 (n=215)		Adults Study 2 (n=212)		Total (n=427)		Children Study 2 (n=162)		Total (n=589)		
	No	%	No	%	No	%	No	%	No	%	
Demographic data											
Sex - Male	127	59.1	120	56.6	247	57.8	69	42.6	316	53.7	P=0.001
Mean age (SD)	44 (20)		44.7 (19.6)		43.1 (19.7)		3.1 (3.1)		36.3 (23.1)		
Prior antibiotic use	33	15.3	71	33.5	104	24.4	45	27.8	149	25.3	P=0.455
Clinical features											
Fever	179	83.3	167	78.8	346	81	151	93.2	497	84.4	P<0.001
Altered consciousness	149	69.3	98	46.2	247	40.7	35	21.6	282	47.9	P<0.001
Headache	153	71.2	118	55.7	271	63.5	57	35.2	328	55.7	P<0.001
Vomiting	116	54.0	58	27.4	174	40.7	84	51.9	258	43.8	P=0.015
Photophobia	53	24.7	39	18.4	92	21.5	10	6.2	102	17.3	P<0.001
Behavioral changes	101	47	53	25	154	36.1	87	53.7	241	40.9	P=0.025
Focal signs	33	15.3	16	7.5	49	11.5	4	2.5	53	9	P=-0.001
Seizures	59	27.4	16	7.5	75	17.6	66	40.7	141	23.9	P<0.001
Meningism	92	42.8	90	42.5	182	42.6	43	26.5	225	38.2	P<0.001

Table 3. Investigations

	Study 1 (n=215) (no./no. performed)		Adults Study 2 (n=212) (no./no. performed)		Total (n=427) (no./no. performed)		Children Study 2 (n=162) (no./no. performed)		Total (n=589) (no./no. performed)	
	No	%	No	%	No	%	No	%	No	%
Blood investigations										
ESR>20	44/140	31.4	96/122	78.7	140/262	53.4	7/8	87.5	147/270	54.4
CRP >12	38/54	70.4	54/105	51.4	92/159	57.9	64/127	50.4	156/286	54.6
WBC>11000	124/201	61.7	100/200	50	224/401	55.9	105/160	65.6	329/570	57.7
Blood Culture positive	1/65	1.5	3/79	3.8	4/144	2.1	5/84	5.9	9/228	3.9
CSF analysis										
CSF done	146	67.9	170	80.2	316	74	162	100	478	81.2
Protein >45	100/146	68.4	105/170	61.8	205/316	64.9	98/162	60.5	303/478	63.4
Sugar<1/2 RBS	28/146	19.2	25/170	14.7	53/316	16.8	24/162	14.8	77/478	16.1
PMN>1	64/146	43.8	47/170	27.7	111/316	35.1	72/162	44.4	183/478	38.3
Lymphocytes>5	72/146	49.3	73/170	42.9	145/316	45.9	90/162	55.5	235/478	49.2
CSF Gram stain positive	0/78	0	0/113	0	0/113	0	0/152	0	0/343	0
CSF culture positive	0/70	0	0/113	0	0/186	0	0/151	0	0/337	0
AFB in CSF positive	0/42	0	0/89	0	0/131	0	0/1	0	0/132	0
Other investigations										
CT scan -										
Cerebral oedema	35/151	23.2	35/165	21.2	70/316	22.2	5/7	71.4	70/323	21.7
EEG Abnormal	19/83	22.9	25/74	33.7	44/157	28	3/18	16.7	47/323	14.6

(Units of measurement: ESR - mm/1st h; CRP - mg/L; WBC - cells/mm³; CSF protein - mg/dL; CSF cell counts - cells/mm³)

Table 4. Diagnosis reached after investigation at CNTH

	Study 1 (n=215)		Adults Study 2 (n=212)		Total (n=427)		Children Study 2 (n=162)		Total (n=589)	
	No	%	No	%	No	%	No	%	No	%
Clinical syndrome										
Meningitis	36	16.7	78	36.8	114	26.7	77	47.5	191	32.4
Encephalitis	33	15.3	39	18.4	72	16.9	4	2.5	76	12.9
Meningo-encephalitis	63	29.3	49	23.1	112	26.2	1	0.6	113	19.2
Cerebral abscess	4	1.9	1	0.5	5	1.1	0	0.0	5	0.8
Uncertain diagnosis	56	26	18	8.5	74	17.3	4	2.5	78	13.2
Alternative diagnosis	23	10.7	27	12.7	50	11.7	76	46.9	126	21.4
Categorization according to diagnostic certainty										
Definitive diagnosis	1	0.5	4	1.9	5	1.2	7	4.3	12	2.0
Probable diagnosis	123	57.2	136	64.7	249	58.5	61	37.7	310	52.6
Possible diagnosis	12	5.6	27	12.7	39	9.1	14	8.6	53	9.0
Uncertain diagnosis	56	26	18	8.5	74	17.3	4	2.5	78	13.2
Alternative diagnosis	23	10.7	27	12.7	50	11.7	76	46.9	126	21.4

Results of the Study 2 Extension – CSF analysis at Oita University, Japan

233 patients in Study 2 (110 adults, 123 children) enrolled during the period July 2009-November 2010 were considered for detailed CSF analysis. CSF samples from 191 patients were sent to the Oita University, Japan. Analysis of these samples confirmed the presence of the following organisms: Dengue virus (1 patient; DEN-2), Echovirus (2 patients; HEcoV 9, HEcoV 25), Adenovirus (7 patients; all HAdV 41). Two patients (out of 81 tested) had immune-mediated encephalitis with anti-NMDAR antibodies³⁹ (Table 6).

In addition to these, two exciting findings were made. Firstly, Human Bocavirus was identified in five

patients by PCR (HBoV-1,2,3). Phylogenetic analysis showed that these genomes shared some similarities with, but were not identical to, those described in Bangladeshi children³⁹. On further analysis of negative CSF samples by deep sequencing, genomes of two new small circular single-stranded DNA (ssDNA) viruses were identified, a Cyclovirus (one patient) and Gemy-circularvirus (three patients). They belong to a group of viruses with circular, replication initiator protein encoding, single stranded DNA (CRESS-DNA) genomes. Phylogenetic analysis revealed that these genomes were distinct from those previously described, and were designated CyCV-SL and GemyCV-SL⁴⁰. The association of bocavirus (HBoV), cycloviruses and gemycircularviruses with meningo-encephalitis had been described only recently^{47,48,49}.

Table 5. Treatment given and Outcome

	Study 1 (n=215)		Adults Study 2 (n=212)		Total (n=427)		Children Study 2 (n=162)		Total (n=589)	
	No	%	No	%	No	%	No	%	No	%
Treatment given (intravenous drugs)										
Penicillin	52	24.2	23	10.8	75	17.6	22	13.5	97	16.5
3 rd generation cephalosporin	181	84.2	170	80.2	351	82.2	112	69.1	463	78.6
Acyclovir	133	61.9	123	58.0	256	59.9	75	46.2	331	56.2
Dexamethasone	82	38.1	62	29.2	144	33.8	44	27.2	188	31.9
Mannitol	66	30.7	38	17.9	104	24.3	27	16.6	131	22.2
Duration of hospital stay (days)										
Mean (SD)	9.2 (6.8)		9.7 (5.2)		9.5 (6.1)		7.1 (3.8)		8.9 (5.7)	
Outcome										
Discharged home	153	71.2	164	77.4	317	74.2	160	98.8	477	81
Death	20	9.3	11	5.2	31	7.26	0	0.0	31	5.3
Transferred for further care	32	14.9	31	14.6	63	14.8	1	0.6	64	10.9
Left hospital – destination unknown	10	4.7	6	2.8	16	3.74	1	0.6	17	2.9

CSF analysis in Japan thus increased the diagnostic yield to 32 out of 374 patients (8.6%) in Study 2. (Table 6) After excluding those with 'uncertain' and 'alternative' diagnoses, the yield was 12.9% among those more likely to have a CNS infection ('definite', 'probable' and 'possible'). However, almost 90% of patients remained without an aetiological diagnosis even after extensive investigation.

Discussion

Key findings and interpretation

This oration summarises the findings of two prospective studies on CNS infections in Sri Lanka. They were conducted in the same setting, over a similar duration, with identical inclusion/ exclusion criteria and similar methodology. They were the first-

ever prospective studies on adult patients in the country, and provide data from the largest case series to date of adult and paediatric Sri Lankan patients.

The key finding from the studies is the difficulty in establishing a microbiological diagnosis, and the resultant low diagnostic yield. In Study 1, a definitive aetiological diagnosis was made in only one out of 215 patients. In Study 2, only 11 out of 374 patients (2.9%) had a definitive diagnosis with the facilities available at the CNTH. On further analysis of CSF at an overseas laboratory, a cause was identified in 21 additional patients, increasing the yield to 32 (8.6%). In about two-thirds (62%) of patients where a diagnosis of CNS infection seemed highly likely ('probable' and 'possible' categories), we could not establish a microbiological causation (Table 4).

Table 6. Aetiological diagnoses identified

Study 1	Study 2 (CNTH analysis)	Study 2 Extension (Analysis in Japan)
<i>Adults</i> (n=215)	<i>Adults + Children</i> (n=374)	<i>Adults + Children</i> (n=191)
Blood culture		
<i>S. pneumoniae</i> (n=1)	<i>S. pneumoniae</i> (n=3) <i>Staph. aureus</i> (n=4) <i>H. influenzae</i> (n=1)	
Serology		
Japanese Encephalitis (n=1)	Enterovirus (n=1)	
CSF PCR		
	<i>H. influenzae</i> (n=1)	Dengue virus – DEN-2 (n=1) Echovirus – HEcoV 9, HEcoV 25 (n=2) Adenovirus – HAdV 41 (n=7) Human Bocavirus – HBoV-1,2,3 (n=5)
Genome detection		
		Cyclovirus – CyCV-SL (n=1) Gemycircularvirus – GemyCV-SL (n=3)
On-cell Western analysis		
		Anti-NMDAR antibody (n=2)
Total aetiologies identified		
n=1, 0.46%	n=11, 2.94%	n=21, 10.9%

Lack of modern diagnostic facilities in the state sector is likely to be the main reason for the very low rate of isolation of organisms. However, suboptimal management practices are possible contributory factors, such as the failure to carry out even the routinely available investigations (e.g, blood cultures were done in only 39%). Delays in performing lumbar puncture and blood cultures, and improper techniques in specimen handling, can lead to negative results. We tried to minimise the influence of these during Study 2 with several measures such as staff briefings. An audit is currently underway to evaluate the management practices in detail with a view to identifying deficiencies and introducing remedial measures. Prior antibiotic use is likely to be an important factor for a poor diagnostic yield, and is common (27-61%) in

developing countries^{27,31,32}, compared to about 15% reported from developed countries³. In our study, 25% of the patients had been treated with antibiotics prior to hospital admission (Table 2).

We were unable to detect any cases of cerebral malaria or TBM. Malaria was almost eradicated in Sri Lanka at the time of the studies. Tuberculosis remains a major health problem in the country, and the failure to detect TBM is likely due to a combination of factors such as inadequate testing (AFB staining, culture) and lack of newer techniques such as TB-PCR. However, our findings also raise concerns regarding the widespread practice of empiric use of anti-TB chemotherapy in suspected CNS infection, based solely on a lymphocytic-predominant CSF without diagnostic confirmation.

Our findings warrant a critical analysis of the available literature. The low isolation rates in our studies are similar to reports from many developing countries, and are usually attributed to the lack of diagnostic facilities^{10,26}. However, low isolation rates are common in developing countries even when supported by good microbiological facilities, as shown by many studies. In two Vietnamese studies, a microbiological cause was not found in 73% of adults⁵⁰, and 59% of children⁵¹. In the South Asian Pneumococcal Alliance Network (SAPNA) study of children aged <5 years with suspected meningitis, bacterial meningitis was confirmed in only 3.1% of CSF samples in Pakistan³², and 8.5% in Sri Lanka (Batuwantudawe et al, 2009)²⁹. In another study of children <5 years in Colombo, an organism was isolated in only 9.4% CSF specimens, and only 2.3% had a positive culture (Batuwantudawe et al, 2010)³⁰. Two Sri Lankan studies, conducted subsequent to our studies, have confirmed the difficulties in diagnosis in spite of access to good microbiological facilities. In a study by Danthanarayana et al (2015), Enterovirus (E-9) was detected in nine out of 39 CSF samples, and the samples were negative for all other viruses tested and bacterial culture⁵². Lohitharahjah et al (2017) identified a viral aetiology in 27.3% of 99 CSF samples from 108 patients with suspected encephalitis/meningo-encephalitis; the aetiologies identified included dengue virus, JE virus, varicella zoster virus and West Nile virus. Bacterial causes and HSV were not detected⁵³.

A closer analysis of the published literature reveals that reported high isolation rates can perhaps be misleading⁵⁴. Most studies reporting high isolation rates are from case series with a defined syndrome (bacterial meningitis, viral encephalitis, etc.), are retrospective studies, or describe only microbiologically or serologically proven cases^{1,2,3,20,55,56}. These would naturally show higher isolation rates. Data from unselected patients with clinically suspected CNS infection, as in our study, is limited^{41-44,57,58}. Such data is more important for 'point of care' clinical decision making in the acute stage, as patients do not present with a label of 'bacterial meningitis' or 'viral encephalitis', but with a diagnostically challenging clinical syndrome of febrile meningo-encephalopathy (meningo-encephalitis)⁵⁴. Similar studies with unselected patient recruitment based on a presumptive clinical diagnosis of CNS infection have reported much lower rates of isolation even from developed countries, similar to our findings^{41-44,58-60}. In a multicentre study from UK, only 20% out of 217 suspected patients were found to have a CNS infection, and CSF culture was positive in only 3 out of 199 samples⁴³. In a national level survey in UK, 60% of the cases were of unknown aetiology⁵⁹. An infectious

cause was detected in only 37% of patients in another multi-centre study in UK⁴⁴. In the California Encephalitis Project, a 'confirmed or probable' cause was found in only 16% of patients, and an aetiological agent was not found in 62% of patients despite extensive testing⁴². Up to 85% of cases worldwide are reported to be due to undetected causes⁴⁴.

It is likely that there are many other potential reasons for a low diagnostic yield. The possibility of a broader spectrum of neurotropic organisms, not considered in the traditional list of differential diagnosis and not detected by the usual tests for CNS infection, needs serious consideration. Furthermore, the epidemiological patterns seem to be changing in South Asia, and worldwide. We found several cases of dengue, echo- and adenovirus infections. Dengue is now considered one of the main aetiologies of encephalitis in endemic regions⁶¹⁻⁶⁴. Increasing global travel can introduce previously unseen organisms to new territories, e.g. West Nile virus encephalitis is now reported from the South Asian region, including Sri Lanka⁶⁵⁻⁶⁷. Emerging infections such as Nipahvirus encephalitis is increasingly recognised from South East Asia⁶⁸⁻⁷¹. Interestingly, the two subsequent Sri Lankan studies cited earlier detected only viral aetiologies and failed to isolate any bacterial causes^{52,53}. Alternate diagnoses such as encephalopathy are common in unselected samples, e.g. 79.7% of suspected CNS infection in a multi-centre study in UK⁴³; 31.3% of suspected bacterial meningitis in Ethiopia⁷²; 10% of suspected encephalitis in the California Encephalitis Project⁴². In our studies, an 'alternate diagnosis' was found in 21.4% (adults - 12.7%, children - 46.9%, $p=0.001$). Furthermore, recent reports indicate that non-infectious causes of encephalitis, such as immune-mediated encephalitis, may be at least as common as infectious causes. In patients aged <30 years in the California Encephalitis Project, immune-mediated encephalitis due to anti-N-methyl-D-aspartate receptor (anti-NMDAR) antibodies was commoner than viral aetiologies⁷³. Immune-mediated encephalitis was found in 21% of patients with encephalitis in England⁴⁴, and in 24% in Thailand⁷⁴. We found two patients with anti-NMDAR encephalitis in Study 2.

Over the years, we may well have been overlooking the possible existence of a completely different spectrum of neurotropic organisms, not previously considered as pathogens causing CNS infection. Several hitherto unsuspected novel agents, such as Bocavirus, Cyclovirus and Gemycircularvirus, were isolated in our patients^{39,40}. The association of Bocavirus with encephalitis was previously described only in one report from Bangladesh⁴⁷. Cyclovirus has

only been previously reported in association with CNS involvement from Vietnamese children and Malawi adults^{48,49}. To our knowledge, ours was the first report of an association of Gemycircularvirus with CNS infection. The causal importance of these organisms in CNS infection is yet to be determined. However, these findings were considered important enough to be highlighted with special mention in the respective journals^{75,76}. Both Bocavirus and Cyclovirus have more recently been reported from patients with encephalitis, increasing their likelihood of being causal pathogens^{77,78}.

Many patients in our studies were treated with intravenous antibiotics on clinical suspicion, without confirmation of CNS infection. This practice is widespread in many developing countries where diagnostic facilities are limited; e.g., bacterial meningitis was confirmed in only 3.3% of patients with suspected meningitis in Ethiopia, but all patients were given intravenous antibiotics⁵⁷, 58% of patients with suspected meningitis in Nigeria were treated empirically without CSF analysis⁷⁹. This is a pragmatic approach to emergent management in view of the high mortality and morbidity associated with CNS infection. However, it also carries the risks of overlooking many potentially treatable diagnoses, and a high cost of unnecessary treatment. Interestingly, no cases of microbiologically confirmed HSV encephalitis were found in our study and in the two subsequent Sri Lankan studies described^{52,53}. Intravenous acyclovir is widely used on an empiric basis in patients with suspected meningo-encephalitis in Sri Lanka, as was seen in our studies, and these findings challenge the rationale for this practice.

Practice guidelines for empiric treatment are based on epidemiological data from developed countries; such data are not available for developing countries. Disease patterns in these countries may well be different, as our findings suggest. Furthermore, there is little evidence for using some of the guideline-based treatments in the developing countries. Adjunctive dexamethasone therapy is widely recommended in bacterial meningitis but several studies have failed to show a benefit in developing countries^{27,28,80}. A study in Ethiopia found excess mortality with adjunctive dexamethasone therapy, but mortality was not increased in the microbiologically proven cases, highlighting the importance of accurate diagnosis guiding empiric treatment⁷².

Strengths and Limitations

The main strengths of these studies include the large number of participants, recruitment of consecutive

patients with suspected CNS infection admitted to all the medical units (and all the paediatric units in Study 2) of a busy teaching hospital, and prospective data collection over a long period. The similarities in the patient numbers, patient characteristics and findings between adults in the two studies point to the goodness of the data.

Several limitations need to be acknowledged. Between-unit variations in practice may have affected diagnostic work up and treatment; we did not investigate this. We also did not investigate the possible impact of deficiencies in management, such as delays in performing investigations. Our findings are from a single urban centre, and data may well be different from other parts of the country. A selection bias is possible as many patients referred from smaller hospitals to a tertiary care centre are included. This data may not be a true reflection of the pattern of CNS infections in the community, as more severe patients may die before reaching hospital and mild undiagnosed cases may be treated at home. However, most patients with acute CNS infections are likely to seek hospital treatment. Community based studies on CNS infections are virtually impossible due to the need for in-hospital evaluation and management, and almost all the reports in the literature are based on hospital derived data. We believe our data is representative of adults and children with CNS infection attending a large Sri Lankan hospital.

Conclusions and Recommendations

These studies highlight the difficulties in establishing a diagnosis in patients with suspected CNS infection in Sri Lankan hospitals. Our findings challenge the traditional approach to the management of CNS infections, and point to the need for 'thinking outside the box' – searching beyond the traditional mind set for a broader range of possible organisms. These findings are strengthened by similar data from other studies, from Sri Lanka and many other countries. Failure to establish an accurate diagnosis would naturally lead to inappropriate treatment, and possibly death and disability. It also has huge financial implications which developing countries such as Sri Lanka can ill afford; e.g., treatment of many patients with expensive antimicrobials on empiric grounds and prolonged hospital stays. Availability of rapid and accurate diagnostic facilities in state sector hospitals is likely to minimise treatment costs, in addition to improving patient care. Investigations such as PCR which are standard practice in other countries should be made available to practicing clinicians.

The challenge for Sri Lankan physicians?

Getting the treatment right in the critical first few hours depends on getting the diagnosis right. This is particularly challenging in resource-limited settings such as Sri Lanka where CNS infections are more prevalent, due to a multitude of factors such as lack of rapid diagnostic facilities, widespread prior antibiotic use and lack of epidemiological data. There is a clear need for national-level research into the aetiology of CNS infections and antibiotic sensitivity patterns. A shift in our approach to CNS infections, and searching beyond the traditional diagnostic paradigms for previously unsuspected infectious agents and non-infectious causes deserves more attention. Use of modern diagnostic tools and newer antibiotics would add to the cost of care, but this would be money well spent, on minimising disability and saving lives.

Acknowledgements

I sincerely thank the following for their contributions: Collaborators at the Faculty of Medicine, University of Kelaniya (EGDS Rajindrajith, KVHKK Perera, BHR Premaratne, Wijesooriya T, HJ de Silva); Collaborators at the Colombo North Teaching Hospital, Ragama (KMMP Dassanayake, SMSB Samarakoon, G Premawansa, K Thirumawalawan, A Kulatunga, MAM Fernando, L de Silva, WAJN Tissera, DKMMS Cooray); Overseas Collaborators (Kamruddin Ahmed, Eric Delwart, D Mori, TG Phan, K Yamada, K Miya, T Matsumoto, MT Mitui, H Mori, A Nishizono, M Söderlund-Venermo, X Deng, TF Fan Ng, F Bucardo-Rivera, P Orlandi); Pre-intern medical officers who collected and entered data (ML Harshani, VNRM Fonseka, KLW Hathagoda, ANHMUKGDB Nawaratne, WALK Weerasooriya, NBANA Kumari, WRS Wimalaratne); Pre-intern medical officers who helped with data analysis (RMSP Muwanhella, DDVLS Bandara); All the staff in medical and paediatric units at the Colombo North Teaching Hospital, Ragama; All the Patients.

References

- Durand ML, Calderwood SB, Weber DJ, et al. Acute bacterial meningitis in adults: a review of 493 episodes. *N Engl J Med*. 1993; **328**: 21-28.
- Køster-Rasmussen R, Korshin A, Meyer CN. Antibiotic treatment delay and outcome in acute bacterial meningitis. *J Infect*. 2008; **57**: 449-54.
- Weisfelt M, van de Beek D, Spanjaard L, et al. Clinical features, complications, and outcome in adults with pneumococcal meningitis: a prospective case series. *Lancet Neurol*. 2006; **5**: 123-9.
- Hosoglu S, Ayaz C, Geyik MF, et al. Tuberculous meningitis in adults: an eleven-year review. *Int J Tuberc Lung Dis*. 1998; **2**: 553-7.
- Farinha NJ, Razali KA, Holzel H, Morgan G, Novelli VM. Tuberculosis of the central nervous system in children: a 20-year survey. *J Infect*. 2000; **41**: 61-8.
- Solomon T, Dung NM, Kneen R, Gainsborough M, Vaughn DW, Khanh VT. Japanese encephalitis. *J Neurol Neurosurg Psychiatry* 2000; **68**: 405-15.
- Solomon T. Flavivirus encephalitis. *N Engl J Med* 2004; **351**: 370-8.
- Bohr V, Paulson OB, Rasmussen N. Pneumococcal meningitis: late neurologic sequelae and features of prognostic impact. *Arch Neurol*. 1984; **41**: 1045-9.
- Baraff LJ, Lee SI, Schriger DL. Outcomes of bacterial meningitis in children: a meta-analysis. *Pediatr Infect Dis J* 1993; **12**: 389-94.
- Scarborough M, Thwaites GE. The diagnosis and management of acute bacterial meningitis in resource-poor settings. *Lancet Neurol*. 2008; **7**: 637-48.
- Duke T, Curtis N, Fuller DG. The management of bacterial meningitis in children. *Expert Opin Pharmacother*. 2003; **4**: 1227-40.
- Wall EC, Cartwright K, Scarborough M, et al. High mortality amongst adolescents and adults with bacterial meningitis in sub-Saharan Africa: an analysis of 715 cases from Malawi. *PLoS One*. 2013; **8**: e69783.
- Gessner BD, Mueller JE, Yaro S. African meningitis belt pneumococcal disease epidemiology indicates a need for an effective serotype 1 containing vaccine, including for older children and adults. *BMC Infect Dis*. 2010; **10**: 22.
- Aronin SI, Peduzzi P, Quagliarello VJ. Community-acquired bacterial meningitis: risk stratification for adverse clinical outcome and effect of antibiotic timing. *Ann Intern Med* 1998; **129**: 862-69.
- Proulx N, Frechette D, Toye B, Chan J, Kravcik S. Delays in the administration of antibiotics are associated with mortality from adult acute bacterial meningitis. *QJM* 2005; **98**: 291-98.
- Miner JR, Heegaard W, Mapes A, Biros M. Presentation, time to antibiotics, and mortality of patients with bacterial meningitis at an urban county medical center. *J Emerg Med* 2001; **21**: 387-92.
- Auburtin M, Wolff M, Charpentier J, et al. Detrimental role of delayed antibiotic administration and penicillin-nonsusceptible strains in adult intensive care unit patients with pneumococcal meningitis: the PNEUMOREA prospective multicenter study. *Crit Care Med*. 2006; **34**: 2758-65.
- Fitch MT, van de Beek D. Emergency diagnosis and treatment of adult meningitis. *Lancet Infect Dis* 2007; **7**: 191-200.
- Attia J, Hatala R, Cook DJ, Wong JG. The rational clinical examination. Does this adult patient have acute meningitis? *JAMA* 1999; **282**: 175-81.

20. van de Beek D, de Gans J, Spanjaard L, Weisfelt M, Reitsma JB, Vermeulen M. Clinical features and prognostic factors in adults with bacterial meningitis. *N Engl J Med*. 2004; **351**: 1849-59.
21. Thomas KE, Hasbun R, Jekel J, Quagliarello VJ. The diagnostic accuracy of Kernig's sign, Brudzinski's sign, and nuchal rigidity in adults with suspected meningitis. *Clin Infect Dis* 2002; **35**: 46-52.
22. Bartt R. Acute bacterial and viral meningitis. *Continuum (Minneapolis)*. 2012; **18**: 1255-70.
23. van de Beek D, de Gans J, Tunkel AR, Wijdicks EF. Community-acquired bacterial meningitis in adults. *N Engl J Med*. 2006; **354**: 44-53.
24. Anon. Bacterial meningitis. *Continuum*. 2002; **8**: 7-26.
25. Zunt JR, Baldwin KJ. Chronic and subacute meningitis. *Continuum (Minneapolis)*. 2012; **18**: 1290-318.
26. Sacchi CT, Fukasawa LO, Gonçalves MG, et al; São Paulo RT-PCR Surveillance Project Team. Incorporation of real-time PCR into routine public health surveillance of culture negative bacterial meningitis in São Paulo, Brazil. *PLoS One*. 2011; **6**: e20675.
27. Nguyen TH, Tran TH, Thwaites G, et al. Dexamethasone in Vietnamese adolescents and adults with bacterial meningitis. *N Engl J Med* 2007; **357**: 2431-40.
28. Scarborough M, Gordon SB, Whitty CJ, et al. Corticosteroids for bacterial meningitis in adults in sub-Saharan Africa. *N Engl J Med* 2007; **357**: 2441-50.
29. Batuwanthudawe R, Karunaratne K, Dassanayake M, et al. Surveillance of invasive pneumococcal disease in Colombo, Sri Lanka. *Clin Infect Dis*. 2009; **48**(2): S136-40.
30. Batuwanthudawe R, Rajapakse L, Somaratne P, Dassanayake M, Abeyasinghe N. Incidence of childhood Haemophilus influenzae type b meningitis in Sri Lanka. *Int J Infect Dis*. 2010; **14**: e372-6.
31. Shah AS, Knoll MD, Sharma PR, et al. Invasive pneumococcal disease in Kanti Children's Hospital, Nepal, as observed by the South Asian Pneumococcal Alliance network. *Clin Infect Dis*. 2009; **48**(2): S123-8.
32. Zaidi AK, Khan H, Lasi R, Mahesar W; Sindh Meningitis Group. Surveillance of pneumococcal meningitis among children in Sindh, southern Pakistan. *Clin Infect Dis*. 2009; **48** (2): S129-35.
33. Stockdale AJ, Weekes MP, Aliyu SH. An audit of acute bacterial meningitis in a large teaching hospital 2005-10. *QJM*. 2011; **104**: 1055-63.
34. Seneviratne R de S, Navasivayam P, Perera S, Wickremasinghe RS. Microbiology of cerebral abscess at the neurosurgical unit of the National Hospital of Sri Lanka. *Ceylon Med J*. 2003; **48**: 14-16.
35. Peiris JS, Amerasinghe FP, Arunagiri CK, et al. Japanese encephalitis in Sri Lanka: comparison of vector and virus ecology in different agro-climatic areas. *Trans R Soc Trop Med Hyg*. 1993; **87**: 541-8.
36. Peiris JS, Amerasinghe FP, Amerasinghe PH, Ratnayake CB, Karunaratne SH, Tsai TF. Japanese encephalitis in Sri Lanka – the study of an epidemic: vector incrimination, porcine infection and human disease. *Trans R Soc Trop Med Hyg*. 1992; **86**: 307-13.
37. Gunawardhana SA, Somaratne SC, Fernando MA, Gunaratne PS. Tuberculous meningitis in adults: a prospective study at a tertiary referral centre in Sri Lanka. *Ceylon Med J*. 2013; **58**: 21-5.
38. Ranawaka UK, Rajindrajith EGDS, Perera KV, Dassanayake KM, Premaratne BA, de Silva HJ. Clinical profile and difficulties in diagnosis of central nervous system infections in adult patients in a tertiary care hospital. *Ceylon Med J*. 2013; **58**: 26-8.
39. Mori D, Ranawaka U, Yamada K, et al. Human bocavirus in patients with encephalitis, Sri Lanka, 2009-2010. *Emerg Infect Dis*. 2013; **19**: 1859-62.
40. Phan TG, Mori D, Deng X, Rajindrajith S, Ranawaka U, Fan Ng TF, Bucardo-Rivera F, Orlandi P, Ahmed K, Delwart E. Small circular single stranded DNA viral genomes in unexplained cases of human encephalitis, diarrhea, and in untreated sewage. *Virology*. 2015; **482**: 98-104.
41. Glaser CA, Honarmand S, Anderson LJ, et al. Beyond viruses: clinical profiles and etiologies associated with encephalitis. *Clin Infect Dis*. 2006; **43**: 1565-77.
42. Glaser CA, Gilliam S, Schnurr D, et al. California Encephalitis Project, 1998-2000. In search of encephalitis etiologies: diagnostic challenges in the California Encephalitis Project, 1998-2000. *Clin Infect Dis*. 2003; **36**: 731-42.
43. Michael BD, Sidhu M, Stoeter D, et al. North West Neurological Infections Network. Acute central nervous system infections in adults- a retrospective cohort study in the NHS North West region. *QJM*. 2010; **103**: 749-58.
44. Granerod J, Ambrose HE, Davies NW, et al. UK Health Protection Agency (HPA). Aetiology of Encephalitis Study Group. Causes of encephalitis and differences in their clinical presentations in England: a multicentre, population-based prospective study. *Lancet Infect Dis*. 2010; **10**: 835-44.
45. Spanos A, Harrell FE Jr, Durack DT. Differential diagnosis of acute meningitis. An analysis of the predictive value of initial observations. *JAMA*. 1989; **262**: 2700-7.
46. Brouwer MC, Tunkel AR, van de Beek D. Epidemiology, diagnosis, and antimicrobial treatment of acute bacterial meningitis. *Clin Microbiol Rev*. 2010; **23**: 467-92.
47. Mitui MT, Tabib SM, Matsumoto T, et al. Detection of human bocavirus in the cerebrospinal fluid of children with encephalitis. *Clin Infect Dis*. 2012; **54**: 964-7.
48. Tan le V, van Doorn HR, Nghia HD, et al. Identification of a new cyclovirus in cerebrospinal fluid of patients with acute central nervous system infections. *MBio*. 2013; **4**: e00231-13.
49. Smits SL, Zijlstra EE, van Hellemond JJ, et al. Novel cyclovirus in human cerebrospinal fluid, Malawi, 2010-2011. *Emerg Infect Dis*. 2013; **19**: 1511-13.
50. Taylor WR, Nguyen K, Nguyen D, et al. The spectrum of central nervous system infections in an adult referral hospital in Hanoi, Vietnam. *PLoS One*. 2012; **7**: e42099.

51. Le VT, Phan TQ, Do QH, et al. Viral etiology of encephalitis in children in southern Vietnam: results of a one-year prospective descriptive study. *PLoS Negl Trop Dis*. 2010; **4**: e854
52. Danthararayana N, Williams DT, Williams SH, Thevanesam V, Speers DJ, Fernando MS. Acute meningoencephalitis associated with echovirus 9 infection in Sri Lanka, 2009. *J Med Virol*. 2015; **87**: 2033-9.
53. Lohitharajah J, Malavige N, Arambepola C, Wanigasinghe J, Gamage R, Gunaratne P, Ratnayake P, Chang T. Viral aetiologies of acute encephalitis in a hospital-based South Asian population. *BMC Infect Dis*. 2017; **17**: 303.
54. Ranawaka UK. The challenge of treating central nervous system infections. *Ceylon Med J*. 2015; **60**: 155-8.
55. Heckenberg SG, de Gans J, Brouwer MC, Weisfelt M, Piet JR, Spanjaard L, van der Ende A, van de Beek D. Clinical features, outcome, and meningococcal genotype in 258 adults with meningococcal meningitis: a prospective cohort study. *Medicine* (Baltimore). 2008; **87**: 185-92.
56. Thigpen MC, Whitney CG, Messonnier NE, et al. Bacterial meningitis in the United States, 1998-2007. *N. Eng. J. Med*. 2011; **364**: 2016-25.
57. Gudina EK, Tesfaye M, Adane A, et al. Challenges of bacterial meningitis case management in Ethiopia. *Trop Med Int Health*. 2016; **21**: 870-8.
58. Mailles A, Stahl JP; Steering Committee and Investigators Group. Infectious encephalitis in France in 2007: a national prospective study. *Clin Infect Dis*. 2009; **49**: 1838-47.
59. Davison, KL, Crowcroft NS, Ramsay ME, Brown DW, Andrews NJ. Viral encephalitis in England, 1989-1998: what did we miss? *Emerg. Infect. Dis*. 2003; **9**: 234-240.
60. Koskiniemi M, Rantalaiho T, Piiparinen H, et al. Infections of the central nervous system of suspected viral origin: a collaborative study from Finland. *J Neurovirol*. 2001; **7**: 400-8.
61. Soares CN, Cabral-Castro MJ, Peralta JM, de Freitas MR, Zalis M, Puccioni-Sohler M. Review of the etiologies of viral meningitis and encephalitis in a dengue endemic region. *J Neurol. Sci*. 2011; **303**: 75-79.
62. Solomon T, Dung NM, Vaughn DW, et al. Neurological manifestations of dengue infection. *Lancet* 2000; **355**: 1053-9.
63. Carod-Artal FJ, Wichmann O, Farrar J, Gascón J. Neurological complications of dengue virus infection. *Lancet Neurol*. 2013; **12**: 906-19.
64. Verma R, Sahu R, Holla V. Neurological manifestations of dengue infection: a review. *J Neurol Sci*. 2014; **346**: 26-34.
65. Anukumar B, Sapkal GN, Tandale BV, Balasubramanian R, Gangale D. West Nile encephalitis outbreak in Kerala, India, 2011. *J Clin Virol*. 2014; **61**: 152-5.
66. Rutvisuttinunt W, Chinnawirotpisan P, Klungthong Cet al. Evidence of West Nile virus infection in Nepal. *BMC Infect Dis*. 2014; **14**: 606.
67. Lohitharajah J, Malavige GN, Chua AJ, Ng ML, Arambepola C, Chang T. Emergence of human West Nile Virus infection in Sri Lanka. *BMC Infect Dis*. 2015; **15**: 305.
68. Chua KB, Goh KJ, Wong KT, et al. Fatal encephalitis due to Nipah virus among pig-farmers in Malaysia. *Lancet* 1999; **354**: 1257-9.
69. Kulkarni DD, Tosh C, Venkatesh G, Senthil Kumar D. Nipah virus infection: current scenario. *Indian J Virol*. 2013; **24**: 398-408.
70. Homaira N, Rahman M, Hossain MJ, et al. Cluster of Nipah virus infection, Kushtia District, Bangladesh, 2007. *PLoS One*. 2010; **5**: e13570.
71. Homaira N, Rahman M, Hossain MJ, et al. Nipah virus outbreak with person-to-person transmission in a district of Bangladesh, 2007. *Epidemiol Infect*. 2010; **138**: 1630-6.
72. Gudina EK, Tesfaye M, Adane A, Lemma K, Shibiru T, Wieser A, Pfister HW, Klein M. Adjunctive dexamethasone therapy in unconfirmed bacterial meningitis in resource limited settings: is it a risk worth taking? *BMC Neurol*. 2016; **16**: 153.
73. Gable MS, Sheriff H, Dalmau J, Tilley DH, Glaser CA. The frequency of autoimmune N-methyl-d-aspartate receptor encephalitis surpasses that of individual viral etiologies in young individuals enrolled in the California encephalitis project. *Clin Infect Dis*. 2012; **54**: 899-904.
74. Saraya A, Mahaviahakanont A, Shuangshoti S, et al. Autoimmune causes of encephalitis syndrome in Thailand: prospective study of 103 patients. *BMC Neurol*. 2013; **13**: 150.
75. Deresinski S. Encephalitis and Bocavirus? *Clin Infect Dis*. 2014; **58**: iii-iv.
76. Anon. Circular DNA viral genomes in cases of encephalitis. Virology Blog - <http://www.virologyhighlights.com/circular-dna-viral-genomes-in-cases-of-encephalitis/>. Posted on June 30, 2015
77. Yu JM, Chen QQ, Hao YX, Yu T, Zeng SZ, Wu XB, Zhang B, Duan ZJ. Identification of human bocaviruses in the cerebrospinal fluid of children hospitalized with encephalitis in China. *J Clin Virol*. 2013; **57**: 374-7.
78. Zhou C, Zhang S, Gong Q, Hao A. A novel gemycircularvirus in an unexplained case of child encephalitis. *Virology*. 2015; **12**: 197.
79. Sanya E, Taiwo S, Azeez O, Oluyombo R. Bacteria Meningitis: Problems Of Empirical Treatment In A Teaching Hospital In The Tropics. *Internet J Infect Dis*. 2006; **6**.
80. Molyneux EM, Walsh AL, Forsyth H, et al. Dexamethasone treatment in childhood bacterial meningitis in Malawi: a randomised controlled trial. *Lancet* 2002; **360**: 211-18.